

RNA-seq: Getting counts

RNA-seq data analysis workflow

(1) Raw gene expression quantification

Trimming

BBDuk

Cutadapt

Trimmomatic

Alignment against genome

HiSat2

STAR

TopHat2

Hybrid alignment (genome + transcriptome)

RUM

Alignment against transcriptome

STAR

Bowtie2

Pseudoalignment

Salmon

Salfish

Kallisto

Counting

Cufflinks

HTSeq*

Stringtie

RSEM

eXpress

Normalization

FPKM

Raw

FPKM

FPKM

FPKM

Effective counts

Estimated counts

TPM

TPM

NumReads

TPM

Estimated counts

(2) Differential gene expression

Ballgown

baySeq

Cuffdiff

DESeq2

EBseq

edgeR exact test**

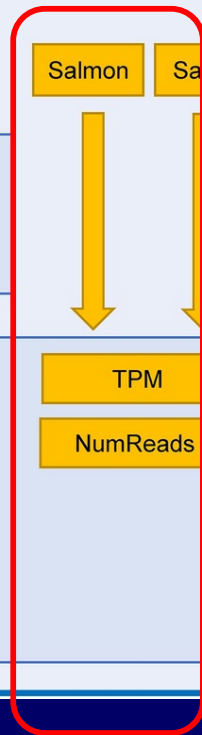
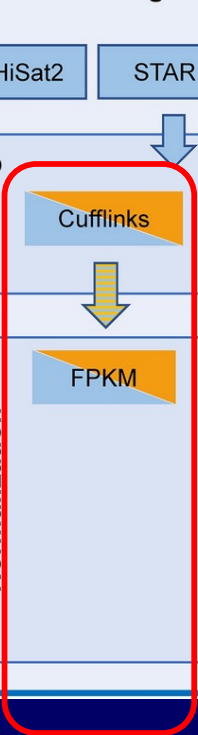
edgeR GLM**

limma trend

limma voom

NOISeq**

SAMseq



RNA-Seq: Getting counts

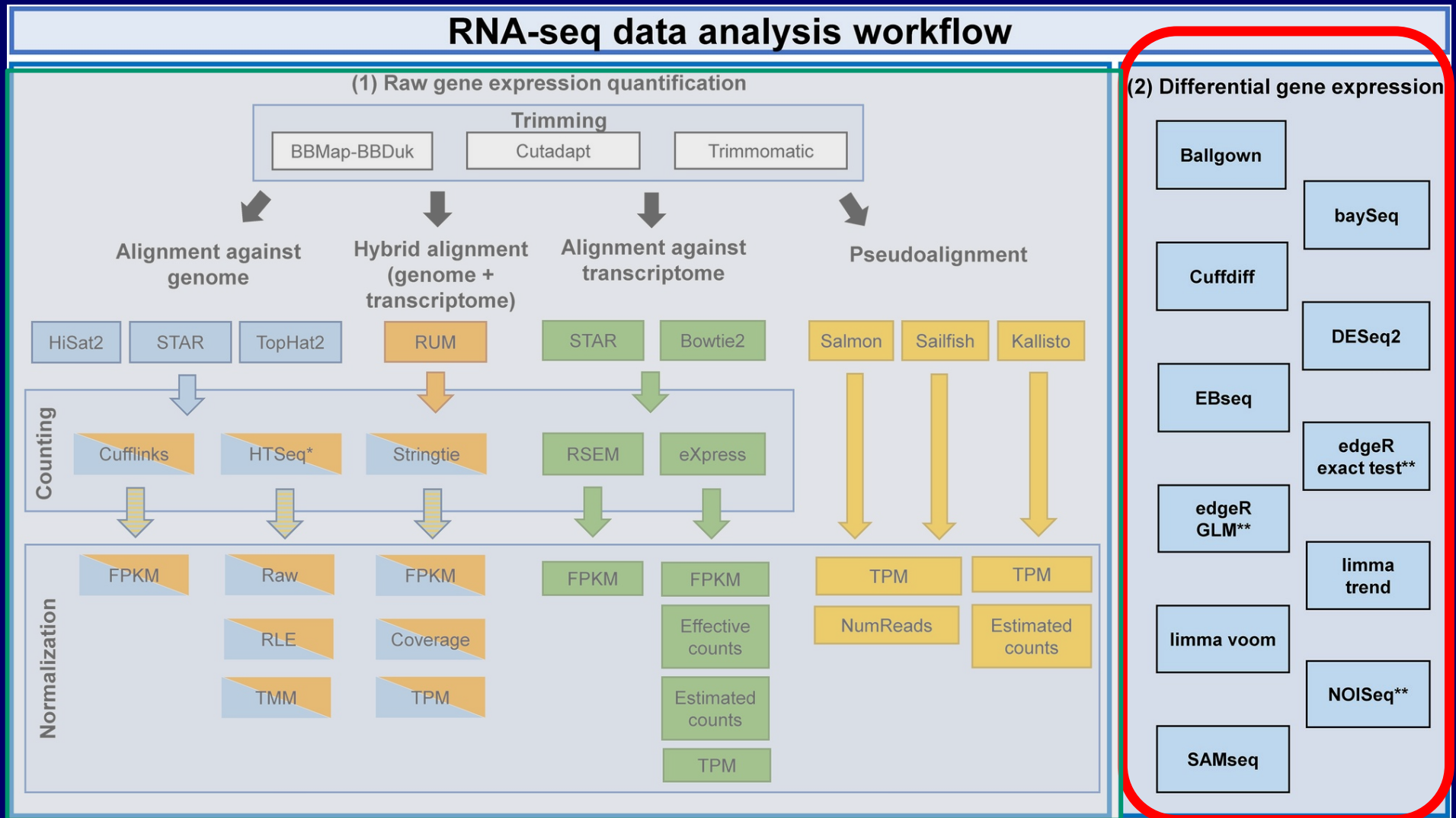
- ❑ Raw – counts (reads) per gene.
- ❑ Normalized
 - ❑ FPKM (Fragments Per Kilobase gene length and per Million reads)
 - ❑ TPM (Transcripts Per Million)
- ❑ Depending on the which program will be used for identifying DEGs.
 - ❑ DESeq (DESeq2) requires raw counts
 - ❑ CuffLinks generated normalized counts as well as models for CuffDiff.

RNA-Seq Overview

Four major steps, **semi-independent** of each other.

- I. Mapping → produce SAM/BAM or counts data.
- II. Quantification → produce RPKM for each gene/transcript.
- III. Identifying DEG (Differentially expressed genes) → gene list.

RNA-seq: Identify DEGs



Many options at this stage. Personal favorites –
Cuffdiff and **DESeq2**

Identification of Differentially Expressed Genes (DEGs)

```
module load cufflinks
```

```
## Frist merge the gtf files for samples to be compared.
```

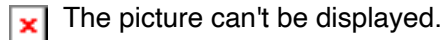
```
In /ufrc/gms6014/share/genome/dm6/annotation/genes.gtf dm6.gtf
```

```
In /ufrc/gms6014/share/genome/dm6/sequence/genome.fa dm6.fa
```

```
cuffmerge -g dm6.gtf -s dm6.fa -p 2 WG_assemblies.txt
```

```
./WG_young_1.clout/transcripts.gtf  
./WG_young_2.clout/transcripts.gtf  
./WG_old_1.clout/transcripts.gtf  
./WG_old_2.clout/transcripts.gtf
```

Representation of (HTS) data – BED (Browser Extensible Data) file



| <u>Chrom.</u> | <u>Start</u> | <u>End</u> | <u>name</u> | <u>Scor</u> | <u>Strand</u> |
|---------------|--------------|------------|-------------|-------------|---------------|
| chr2 | 10000192 | 10000217 | U0 | 0 | + |
| chr2 | 10000227 | 10000252 | U1 | 0 | - |
| chr2 | 10000310 | 10000335 | U2 | 0 | + |
| chr3 | 10000496 | 10000521 | U1 | 0 | - |
| chr2 | 10000556 | 10000581 | U2 | 0 | + |

With
nee

same as the reference genome).

Detailed description of genomic data formats:

<http://genome.ucsc.edu/FAQ/FAQformat.html>

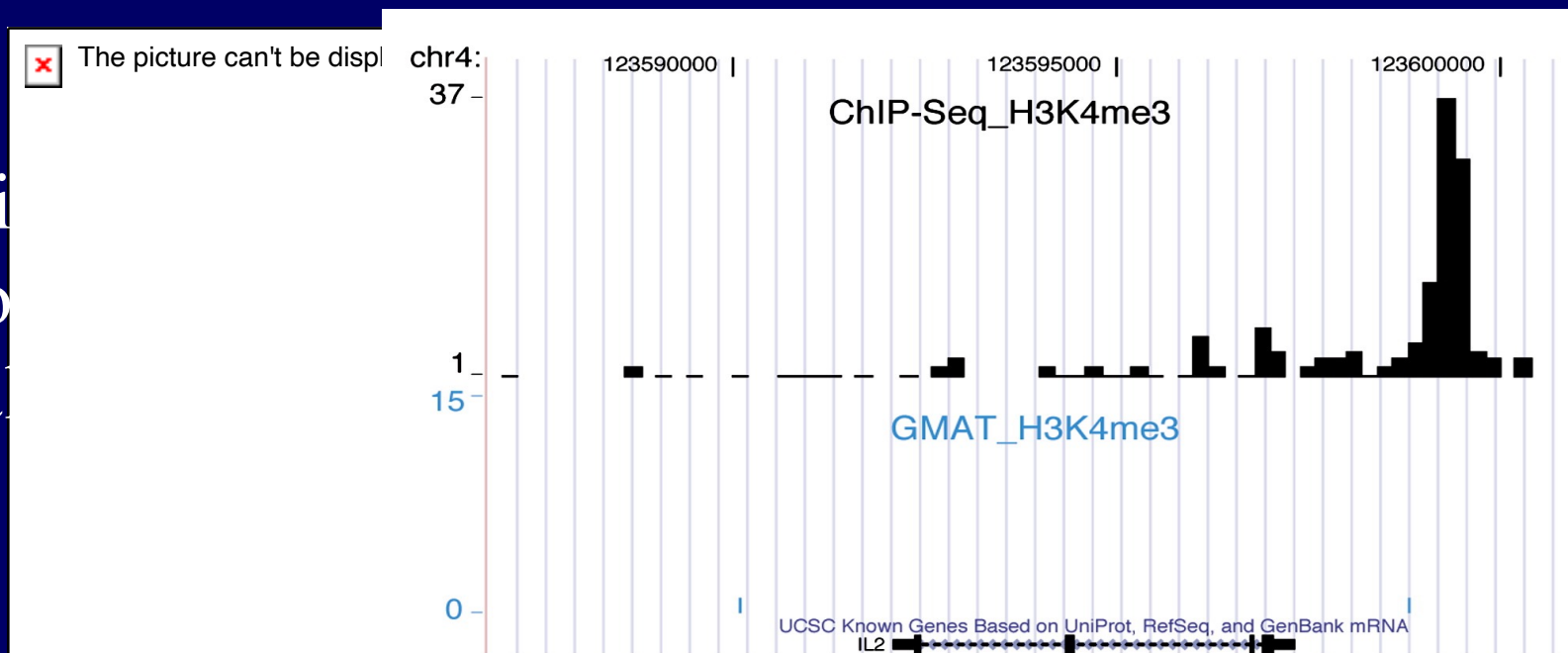
o
e

Representation of (HTS) data – Wig file

```
track type=wiggle_0 name="S_P53_XR60_A_treat_all"  
description="P53_XR"variableStep chrom=chr2L span=10  
11      2  
21     3  
31     4  
41     4  
51     5  
61     4  
71     4  
81     3  
91     3  
101    2  
111    1
```

Visualization of HTS data.

Simple
visualization
distribution
(or normal
values).



Barski et al.
(2007) Cell

| <u>Chr.</u> | <u>ChrStart</u> | <u>ChrEnd</u> | <u>Value</u> |
|-------------|-----------------|---------------|--------------|
| chr4 | 0 | 200 | 0 |
| chr4 | 200 | 400 | 2 |
| chr4 | 400 | 600 | 13 |
| chr4 | 600 | 800 | 35 |
| chr4 | 800 | 1000 | 27 |

BedGraph file (Wig)

Visualizing Deep Seq data with UCSC genome browser

Practice & Observe I:

1. Load the track file as custom track to the browser by copy/past the URL link or upload the file.
2. View 'dense' and then 'full' presentation of the track.

Identification of differentially expressed genes (DEGs)

```
module load cufflinks
```

```
cuffdiff -o Old_v_Young -b ./index/Dm6.44.fa -u Merged/merged.gtf -p 2 -L youngWG,oldWG \  
./starMap/WG_young_1Aligned.sortedByCoord.out.bam,./starMap/WG_young_2Aligned.sortedByCoord.out.bam \  
./starMap/WG_old_1Aligned.sortedByCoord.out.bam,./starMap/WG_old_2Aligned.sortedByCoord.out.bam
```